

UNIVERSIDADE FEDERAL DO RIO GRANDE DO NORTE

CENTRO DE CIÊNCIAS DA SAÚDE CURSO DE GRADUAÇÃO EM FARMÁCIA

INGRID DA SILVA ALBUQUERQUE

SELECTION OF SUITABLE POLYMER AND OBTAINING NEW AMORPHOUS SOLID DISPERSIONS WITH FERULIC ACID AND SOLUPLUS®

NATAL, RN

2020

SELECTION OF SUITABLE POLYMER AND OBTAINING NEW AMORPHOUS SOLID DISPERSIONS WITH FERULIC ACID AND SOLUPLUS®

Trabalho de Conclusão de Curso apresentado ao Curso de Graduação em Farmácia da Universidade Federal do Rio Grande do Norte, como requisito parcial para obtenção do título de Bacharel em Farmácia.

Orientador: Prof. Dr. Ádley Antonini Neves de Lima

Co-Orientadora: Fernanda Ílary Costa Duarte

Universidade Federal do Rio Grande do Norte - UFRN Sistema de Bibliotecas - SISBI Catalogação de Publicação na Fonte. UFRN - Biblioteca Setorial do Centro Ciências da Saúde - CCS

Albuquerque, Ingrid da Silva. Selection of suitable polymer and obtaining new amorphous solid dispersions with ferulic acid and soluplus / Ingrid da Silva Albuquerque. - 2020. 39f.: il. Trabalho de Conclusão de Curso - TCC (Graduação em Farmácia) -Universidade Federal do Rio Grande do Norte, Centro de Ciências da Saúde, Departamento de Farmácia. Natal, RN, 2020. Orientador: Ádley Antonini Neves de Lima. Coorientadora: Fernanda Ílary Costa Duarte.
1. Solubilidade - TCC. 2. Ácido ferúlico - TCC. 3. Dispersões Sólidas Amorfas - TCC. 4. Soluplus® - TCC. I. Lima, Ádley Antonini Neves de. II. Duarte, Fernanda Ílary Costa. III. Título. RN/UF/BS-CCS CDU 544.351.3

INGRID DA SILVAALBUQUERQUE

SELECTION OF SUITABLE POLYMER AND OBTAINING NEW AMORPHOUS SOLID DISPERSIONS WITH FERULIC ACID AND SOLUPLUS®

Trabalho de Conclusão de Curso apresentado ao Curso de Graduação em Farmácia da Universidade Federal do Rio Grande do Norte, como requisito parcial para obtenção do título de Bacharel em Farmácia.

Orientador: Prof. Dr. Ádley Antonini Neves de Lima

Co-Orientadora: Fernanda Ílary Costa Duarte

Presidente: Prof. Ádley Antonini Neves de Lima, Dr.- Orientador, UFRN



Membro: Verônica da Silva Oliveira, Dra., UFRN

NATAL-RN, 18/ junho/2020

AGRADECIMENTOS

Primeiramente, agradeço a Deus, por em todos os momentos e meus caminhos, foi Ele quem me guiou e me deu forças para lutar cada batalha durante minha vida e academia. Obrigada a minha mãe, Maria Cristina, por toda ajuda, cuidado, confiança e paciência durante toda minha trajetória acadêmica, pois sabemos que não foram dias fáceis e que com toda certeza, sem você ao meu lado dia após dia, a realização desse sonho não teria sido possível. Acredito que nunca serei grata ao suficiente por tudo que fez por mim. Agradeço também ao meu pai, Ismael, por sempre ter acredito em mim e ter sido compreensível por todas as faltas e dias sem lhe ver por conta das exigências e obrigações que a mim eram dadas, mas com a certeza que no final a vitória chegaria.

Agradecimento em especial ao meu orientador, Dr. Ádley Antonini, que por meio de um simples e-mail enviado, me aceitou e me acolheu no seu grupo, depositando em mim toda confiança. Obrigada professor por toda recepção, ensinamentos, oportunidades e por ter acreditado no meu potencial desde o princípio.

Um imenso obrigada a minha co-orientadora, Fernanda Ílary, por todo apoio, paciência, dias em laboratórios e de estudos – mesmo com toda dificuldade diante da sua rotina, pelos diversos ensinamentos não somente técnicos mas para a vida e por ter acreditado em mim. Com toda certeza, você foi peça essencial para o meu crescimento na ciência e na vida, serei eternamente grata e nunca esquecerei de tudo que fizestes por mim. Obrigada a todos do grupo INOFARM, pela acolhida, pelo carinho e por todas ajudas durante os anos de pesquisa, com certeza, construí amizades que levarei para sempre.

Gratidão a todos meus amigos que acreditaram e torceram por mim durante toda a faculdade, em especial as minhas amigas Heloísa e Thalia, as quais sempre estiveram ao meu lado nos dias bons e ruins durante esses quatro anos e meio. Agradeço a vocês eternamente por todos os momentos partilhados e que com certeza serão para sempre minhas eternas irmãs que Deus me deu.

Agradeço a todos professores, técnicos e auxiliares de limpeza da faculdade que fizeram com que minha formação fosse possível.

Obrigada a todos vocês que diretamente e indiretamente fizeram parte desse sonho!

"Tudo posso naquele que me fortalece" Filipenses 4:13

RESUMO

Ácido ferúlico (FA) é uma substância com atividades promissoras, entretanto, sua solubilidade aquosa limitada prejudica a obtenção de novas formulações. Diversas estratégias tecnológicas têm sido desenvolvidas para incrementar a solubilidade de fármacos, dentre essas tem-se as dispersões sólidas amorfas (ASD). Assim, o objetivo do presente estudo foi desenvolver uma ASD contendo FA utilizando diferentes métodos de obtenção. O Soluplus® foi selecionado por meio de estudo de solubilidade com o FA, a partir disto foram obtidas ASD com ácido ferúlico por mistura física (PM), malaxagem (KND) e rota evaporação (ER). As dispersões sólidas foram quantificadas por espectrofotometria UV/Vis e caracterizadas físico quimicamente por difração de Raio X (XRD), espectroscopia de infravermelho com transformada de Fourier (FTIR), microscopia eletrônica de varredura (SEM) e calorimetria exploratória diferencial (DSC). Os resultados demonstraram a formação de dispersão solida amorfa através das técnicas utilizadas, sendo que KND e ER demonstraram melhores resultados com significativas reduções das intensidades das reflexões cristalinas pelo XRD, como também, na intensidade das bandas no FTIR e a ausência do pico de fusão característico do FA no DSC das amostras. Dessa maneira, pode-se concluir que foram obtidas dispersões sólidas amorfas com ácido ferúlico que serão investigadas posteriormente quanto à dissolução e atividade antioxidante in vitro.

Palavras-chave: Ácido Ferúlico, Dispersões Sólidas Amorfas, Soluplus®

ABSTRACT

Ferulic acid (FA) is a substance with promising activities, however, its limited aqueous solubility impairs the obtaining of new formulations. Several technological strategies have been developed to increase the solubility of drugs, among which there are amorphous solid dispersions (ASD). Thus, the aim of the present study was to develop an ASD containing FA using different obtaining methods. Soluplus® was selected by solubility study with FA, from this were obtained ASD with ferulic acid by physical mixture (PM), kneading (KND) and evaporation rotary (ER). The solid dispersions were quantified by UV/Vis spectrophotometry and characterized physical-chemically by X-ray diffraction (XRD), Fourier transformed infrared spectroscopy (FTIR), scanning electron microscopy (SEM) and differential exploratory calorimetry (DSC). The results demonstrated the formation of amorphous solid dispersion through the techniques used, and KND and ER showed better results with significant reductions in the intensities of crystalline reflections by the XRD, as well as in the intensity of the bands in the FTIR and the absence of the characteristic fusion peak of FA in the DSC of the samples. Thus, it can be concluded that solid dispersions with ferulic acid were obtained, which will be investigated later for dissolution and antioxidant activity in vitro.

LIST OF FIGURES

Figure 1. Chemical structure of ferulic acid

Figure 2. FA solubility diagram with polymers performed in triplicate

Figure 3. Absorption spectrum of ferulic acid in the UV/Vis region

Figure 4. Spectrum in the UV region of ferulic acid and Soluplus®

Figure 5. FA analytical curve at concentrations from 2 to 12 μ g/mL obtained by UV/Vis spectrophotometer

Figure 6. FA diffractograms, Soluplus®, PM, KND and ER methods

Figure 7. FTIR Spectra for FA, Soluplus®, PM, KND and ER methods

Figure 8. SEM of FA (A), PM (B), KND (C) and ER (D) with increase in 500x

Figure 9. Calorimetric curves of FA, Soluplus®, PM, KND and ER methods

LIST OF TABLES

- **Table 1.** Gibbs free energy for each polymer concentration
- **Table 2**. Absorbance values obtained in the construction of the analytical curve of FA
- **Table 3**. Values obtained for repeatability accuracy
- **Table 4**. Values obtained for intra-day accuracy
- **Table 5**. Accuracy values at three levels of FA concentration.
- **Table 6.** FA content in solid dispersions and efficiency of incorporation (%)

	1.	Introc	luction	•••••					11
	2.	Mater	rials and m	nethods					13
							2.1	Materials	13
							2.2	Methods	13
2.2.1		I	Phase		solubility	diagram	with	polymers	
			•••••						. 13
		2.2.1.	1 Gibbs F	ree Energy	Determination		,		. 13
2.2.2			Preparatio	on of solid d	ispersions			14	
2.2.3			Yield of s	olid dispers	ions			14	
2.2.4			Quantifica	ation of soli	d dispersions			14	
		2.2.5	Incorporat	tion efficien	ncy (IE)				. 15
		2.2.6	Validation	n of the anal	lytical method for	or quantification of	of ferulio	e acid in sol	id
		disper	rsions						. 15
		2.2.7	Characteri	ization of sc	olid dispersions				. 18
	3. R	Results	and discus	ssion					19
	3	.1 Stud	y of feruli	c acid solub	oility in the prese	ence of polymers.			. 19
	3	.2 Vali	dation of t	the analytica	al method by UV	V/VIS spectrophot	ometry		. 21
		3.2.1	Spectral so	canning			, 		. 21
		3.2.2	Selectivity	У					. 21
		3.2.3	Linearity						. 22
		3.2.4	Precision						. 23
		3.2.5	Accuracy						. 24
		3.2.6	Detection	and Quantiz	fication Limits				. 25
	3	.3	Qu	antification	of solid dispers	ions			25
	3	.4	Ch	emical phys	sical characteriza	ation			26
		3.4.1	X-ray diff	raction (XR	2D)				. 26
		3.4.2	Fourier tra	ansform infi	rared spectroscoj	py (FTIR)			. 27
		3.4.3.	Scanning	electron mi	icroscopy (SEM))			. 29
		3.4.4.	Different	ial Explorat	ory Calorimetry				. 30
		4. Co	onclusion.						. 33
	5.	Refer	ences						34

SUMMARY

1. Introduction

In recent times, several Active Pharmaceutical Ingredients (APIs) have been developed, however, the vast majority of these new products have low solubility in water, which is an obstacle in the development of new formulations, especially solid ones for oral administration, such as tablets and capsules. These formulations are usually more stable, with greater ease in their production, have a correct dose and are simpler for administration, as well as have greater adherence when compared to the others¹.

It is known that the solubility of APIs is directly correlated with their bioavailability, for this the Biopharmaceutical Classification System (BCS) establishes an order of classification of drugs according to their absorption. Compounds with low aqueous solubility are inserted in Class II (high permeability, low solubility) or Class IV (low permeability, low solubility)². Studies reveal that about 75% of the compounds under development fall into the Class II and Class IV groups, in addition, of the existing drugs approximately 40% also have low aqueous solubility, so this property is configured as an obstacle in the promotion of new pharmaceutical products^{3,4}. Therefore, in search of improvements and new formulations, scientists and industries are in constant search for new technologies aimed at overcoming these challenges, for example there is the use of salts, development of cocrystals, complexation with cyclodextrins, polymorphs, solid dispersions and particle size reduction⁵.

Amorphous solid dispersions (ASD) are currently established as a platform technology for the formulation of few soluble drugs. The solid dispersion system (SD) consists of a hydrophobic drug dispersed in a hydrophilic vehicle, which will classify SD in first class (sugars and urea), second class (synthetic or natural polymers) and third class (surfactants, mixtures surfactants or polymer mixture)⁶. In addition, the main objective of the ASD is to minimize the packaging energy of the crystal, interrupting its crystalline structure, resulting in an amorphous material that together with the polymer network corroborates the increase in solubility, consequently, the dissolution rate, since hydrophilic transporters can reduce agglomeration and promote a release of the drug in a state of supersaturation, as a consequence increases absorption and improved bioavailability⁷. This generation of a supersaturated state and inhibition of precipitation has been referred to as the "spring and parachute" effect, and this effect is dependent on the type and molecular weight of the polymer⁸.

Ferulic acid (FA) or chemically known as 4-hydroxy-3-metoxicinamic acid (**Figure 1**) is found in abundance in the plant kingdom which has a low toxicity, can be found in whole grains, spinach, grapes, cereal seeds such as wheat, oats and rye, in addition, corn and wheat bran, artichoke and beetroot^{9,10}. This phenolic compound through its structural portions of its molecule is able to combat reactive oxygen and nitrogen species, as well as promote a down regulation in the responses of the pathways of cell death, such as activating the synthesis of proteins that improve cellular stress¹¹. Therefore, the main therapeutic activities originated from FA are due to its antioxidant and anti-inflammatory properties, but also has biological effects such as antibacterial, antifungal, antidiabetic, anti-aging, radioprotection a and anticarcinogenic¹². Also, in this context, it is known that FA is absorbed by the small intestine, is excreted by the renal route and that for pharmacological efficacy will depend on physiological concentrations as well as pharmacokinetic activities, such as absorption¹³.



Figure 1. Chemical structure of ferulic acid

Despite the diversity of biological properties and activities, FA has limited therapeutic application because it presents low aqueous solubility, 6.63 mg.dL⁻¹ at pH 7.2, as well as demonstrating instability of the molecule which can decompose in inactive products, induced by light and/or oxidation¹⁴. Although the chemical structure presents polar groups, the presence of the phenolic group may compromise the solubility in water of this compound, considering that even being able to promote hydrogen bonds with water, there is a high number of carbons in the chain corroborating to reduce solubility, in addition, the presence of the ether group also contributes to this molecule property¹⁵. Therefore, some technological methods have already been developed in order to improve these limiting characteristics, such as inclusion complexes with cyclodextrins and cocrystals¹⁶.

Given the need to optimize the therapeutic potential of ferulic acid, in order to preserve its biological activities and increase its bioavailability, it becomes relevant the use of obtains capable of stabilizing it. In this context, the present study aimed to develop amorphous solid dispersions elaborated by kneading method and evaporation rotary, to characterize the physicochemical properties of these systems.

2. Materials and methods

2.1 Materials

Ferulic acid was purchased from FAGRON® (Lot: 18F23-BO17-0348) and the polymers used were hydroxypropylmethylcellulose (HPMC), polyvinyl caprolactamacetate polyvinyl-polyethylene glycol (Soloplus®), vinyl copolymer pyrrolrolidonevinyl acetate (Kollidon® VA64), polyvinylpyrrolidone (PVP K30) and polyethylene glycol 6000 (PEG 6000); All solvents used are analytical grade.

2.2 Methods

2.2.1 Phase solubility diagram with polymers

10 mL of aqueous solution (using Milli-Q water) of the polymers was added at increasing concentrations (0%; 0.1%; 0.2%; 0.4%; 0.6%; 0.8%; 1.0%). Specifically for the HPMC due to its lower solubility in water, the concentrations tested were 0%; 0.1%; 0.2%; 0.4%.

The solutions were under agitation for 24 hrs, protected from light and after this procedure they were filtered and analyzed in UHPLC-DAD, using a previously validated method¹⁷.

2.2.1.1 Gibbs Free Energy Determination

Gibbs free energy values were used to calculate the spontaneity of the drug solubilization process in aqueous solutions.

From the equation:

$$\Delta G = -2.303RT. \log \frac{Sc}{So}$$
(Equation 1)

Where:

R: Universal constant of perfect gases (8.314472 J.K⁻¹.mol⁻¹);

T : Temperature in Kelvin;

Sc: FA solubility at a given carrier concentration and;

So: concentration of FA in the absence of polymer.

2.2.2 Preparation of solid dispersions

2.2.2.1 Phisycal mixture (PM)

The FA and the respective polymer sat in the solubility study were weighed in proportion (1:1 w/w) and homogenized with pestle and mortar. Subsequently stored in a dissector and protected from light.

2.2.2.2 Kneading method (KND)

The FA and the respective polymer scans selected in the solubility study were weighed in proportion (1:1 w/w) and homogenized with pestle and mortar, then 10% of the total weight volume was added to the ethanol solution: water (50:50 v/v) until the formation of a moist mass, as paste. It was taken to the greenhouse for approximately 24h at 50°C \pm 2°C. Subsequently stored in a dissector and protected from light.

2.2.2.3 Evaporation Rotary method (ER)

The FA and its polymers were weighed in proportion (1:1 p/w) and dissolved in an ethanol:water solution (50:50 v/v) followed by vacuum evaporation at $45^{\circ}C \pm 1^{\circ}C$ using the 150rpm rotaevaporator until complete drying. Subsequently stored in a dissector and protected from light.

2.2.3 Yield of solid dispersions

The yield of the solid dispersions obtained was calculated by means of the ratio between the final mass after preparation, by the initial mass of FA plus polymer, depending on the

Following equation:

$$Yield(\%) = \frac{final mass(SD)}{initial mass(FA+polymer)} x 100$$
(Equation 2).

2.2.4 Quantification of solid dispersions

The quantification of FA present in the solid dispersions (mg/g) developed was performed by spectrophotometry in the ultraviolet region, in evolution 300 UV-Vis spectrophotometer with wavelength analysis at 311 nm, by previously validated method,

using quartz buckets.

Solutions were prepared with solid dispersions in mass equivalent to 5mg of ferulic acid, being diluted in Milli-Q water, obtaining a theoretical concentration of 100μ g/mL. This material was sonicated for 30 min and after proper dilution the UV reading was performed, and quantification was performed in triplicate, in relation to the previously validated method. All the solutions were protected from light.

2.2.5 Incorporation efficiency (IE)

Considering the results obtained by the quantification of ferulic acid, equation 3 was used to obtain the efficiency of incorporation based on the masses of initially added FA and the value of this, present in the solid dispersions obtained¹⁸.

$$IE = \frac{FA \text{ mass in solid dispersions}}{\text{initial mass of } FA} X \text{ 100 (Equation 3).}$$

2.2.6 Validation of the analytical method for quantification of ferulic acid in solid dispersions

The validation of the analytical method was performed according to the criteria of the Resolution of the Collegiate Board (RDC) no. 166 of July 25, 2017 of the Agência Nacional de Vigilância Sanitária (ANVISA)¹⁹.

The parameters analyzed were spectral scanning, selectivity, linearity, intermediate precision and repeatability, accuracy, detection limit and limit of quantification and the tests were performed on Spectrophotometer Evolution 300 UV-Vis, using quartz buckets with 1cm of optical path and wavelength at 311 nm.

2.2.6.1 Stock Solution Preparation

The standard solution containing FA was prepared by solubilizing 10 mg of FA in 100mL of Milli-Q water, obtaining a concentration of 10 μ g/mL and for other analyses at different concentrations, the same solvent was used for dilution. These solutions were protected from light for the achievement of other analyses.

2.2.6.2 Spectral Scan

To perform the scan, a pure FA solution was prepared at a concentration of

 20μ g/mL and a reading of 200 to 600 nm was performed, analyzing the peak of highest absorption.

2.2.6.3 Selectivity

To perform the selectivity, a pure FA solution was prepared at a concentration of $20\mu g/mL$ and another solution containing Soluplus® at 200 $\mu g/mL$, and dilutions were performed necessary to read and obtain the spectral graph.

2.2.6.4 Linearity

For linearity study, three analytical curves were elaborated with seven concentration levels (2,4,5,6,8,10,12 μ g/mL), starting from a stock solution of 100 μ g/mL. These concentrations followed an interval of 80% to 120% of the test concentration (5 μ g/mL), with three replicates for each concentration level. The equation of the line was determined by the mean of the three analytical curves, plotted the values of the concentration as a function of absorbance, thus obtaining the angular coefficient of the line, point of intersection and correlation coefficient (R2), having as acceptance criterion R2 greater than or equal 0.99.

2.2.6.5 Precision

This parameter was evaluated from stock solutions at 100 μ g/mL, performing determinations of three levels, being a high (12 μ g/mL), medium (5 μ g/mL) and low (2 μ g/mL). The precisions were determined by the RSD (relative standard deviation) of the actual concentrations found and by the statistical test ANOVA.

2.2.6.5.1 Repeatability accuracy

The determination of accuracy by repeatability was performed from three stock solutions (100 μ g/mL) analyzing in triplicate the three concentrations being one high (12 μ g/mL), medium (5 μ g/mL) and low (2 μ g/mL), totaling nine determinations. This analysis occurred by the same analyst and equipment.

2.2.6.5.2 Intermediate accuracy

For this criterion, the agreement between the results obtained in the same laboratory under the same conditions was evaluated, however, being performed by analysts and on different days. In total, nine determinations of three concentrations were made, one high (12 μ g/mL), medium (5 μ g/mL) and low (2 μ g/mL) from three stock solutions of 100 μ g/mL.

2.2.6.6 Accuracy

The accuracy of the method was verified from nine determinations within the linear range, three concentrations with three replicates each. Therefore, low concentration solutions (2 μ g/mL) were analyzed. average (5 μ g/mL) and high (12 μ g/mL), according to the linearity limits defined above.

Accuracy was expressed by the ratio between the average concentration determined and the theoretical concentration, according to the formula below:

$$A(\%) = \frac{OME}{CMT} X 100 (Equation 4)$$

Where:

A (%): accuracy in percentage.

CME: experimental mean concentration.

CMT: theoretical mean concentration.

2.2.6.7 Detection and quantification limit

The detection limit (DL) and quantification limit (QL) were calculated based on the relationship between the standard deviation of the analytical curve and its inclination, using the multiplier factors according to RDC $n^{\circ}166$, according to Equations 5 and 6.

$$DL = \frac{3,3 \times SD}{CI}$$
 (Equation 5)

$$QL = \frac{10 \ x \ SD}{CI}$$
 (Equation 6)

Where:

CI: curve slope;

SD: standard deviation.

2.2.7 Characterization of solid dispersions

2.2.7.1 X-ray diffraction (XRD)

The crystalline characterization and crystalline profile of the dispersions were obtained in a Bruker diffratometer, model D2 Phaser, using CuK α radiation (λ =1.54Å) with a Ni filter, in the variation of 10-70° (2 θ), with a step of 0.02°, current of 10 mA, voltage of 30kV, using a Lynxeye detector.

2.2.7.2 Fourier transformed infrared spectroscopy (FTIR)

The isolated components and dispersions obtained were characterized by FTIR, using shimadzu spectrometer, model IR Prestige-21. The spectra were obtained with spectral resolution of 4 cm⁻¹ and 20 scans in the region of 400 to 4000 cm⁻¹.

2.2.7.3 Scanning Electronic Microscopy (SEM)

The samples were metallized with gold and then mounted on aluminum stubs using double-sided tape. Subsequently, they were analyzed by high-resolution scanning electron microscopy. Secondary electronic images were acquired with an acceleration voltage between 5 and 10 kV. Images of retrodiffusion electrons being performed with an acceleration voltage of 15 kV.

2.2.7.4 Differential Exploratory Calorimetry (DSC)

Differential Exploratory Calorimetry (DSC) was performed in Shimadzu® Model DSC-50, weighing approximately 2.0 mg of the sample in hermetically sealed aluminum crucible, under nitrogen atmosphere, purge gas flow of 100 mL.min⁻¹, heating ratio of 10° C.min⁻¹ with final temperature of 250°C. The analysis of the curves obtained was performed using TA Analysis software.

3. Results and discussion

3.1 Study of ferulic acid solubility in the presence of polymers

The objective of this analysis is to observe the influence of polymers on the solubility of ferulic acid according to the increase in the concentration of carriers in the solutions. These results can be analyzed according to the figure obtained through studies conducted in UHPLC, in which it is important to verify the increasing linearity of the curves that each polymer presented according to the concentration used and the percentage of solubilization for ferulic acid in solution.

According to figure 2 it can be perceived that the copolymer Soluplus® was the best polymer increasing the solubility of FA from 15.40 to 36.45 μ g/mL at the concentration of 1%, corroborating the result of Shamma and Basha²⁰, which obtained better solubility of Carvedilol in the presence of Soluplus®. It is also worth emphasizing the importance of evaluating Gibbs free energy on these prepared solutions, correlating spontaneity in the solubilization process. In general, it is known that when $\Delta G^{\circ} < 0$, the greater the increase in solubility by polymers according to the increase of their concentrations, therefore, the more negative the result, the better the solubilizer effect of the carriers. According ly, it can be seen that Soluplus® also presented the most negative Gibbs free energy at the concentration of 1%, and that it was proportional to the increase in concentration, as shown in Table 1.

Soluplus® is a polyvinyl caprolactam copolymer - polyvinyl acetate - polyethylene glycol, classified as a member of the fourth generation of carriers for SDs, which aims to achieve the highest degree of increase in the dissolution of drugs with low aqueous solubility. Soluplus® is a polymeric solubilizer with amphiphilic structure, with a large number of groups with hydroxyls present, making it a good carrier^{21,22}.

Therefore, according to this result and the characteristics of Soluplus®, in order to develop a good solid dispersion, this polymer was chosen for the development of the formulations, as well as the characterizations performed in this work.



Figure 2. Curves of the FA solubility diagram with polymers performed in triplicate

CONCENTRATION	PVPK30	KOLLIDON	SOLUPLUS	PEG	HPMC
(%)					
0.1	-686.37	-590.78	-701.77	-39.96	14.50
0.2	-827.07	-590.78	-701.77	-39.96	14.50
0.4	1209.07	902 52	1610 44	00 07	96 17
0.4	-1208.97	-093.32	-1010.44	-90.07	00.17
0.6	-1280.2	-1149.66	-2106.6	-199.06	
0.8	-1371.41	-1560.53	-2298.09	-248.98	
1.0	-1500.18	-1712.43	-2432.47	-370.83	

Table 1. Gibbs free energy for each polymer concentration

3.2 Validation of the analytical method by UV/VIS spectrophotometry

3.2.1 Spectral scanning

To start the validation protocol, the scan was performed in order to identify the spectral profile of FA in the UV-Vis region. Thus, it was found that the maximum absorption occurs at a wavelength of 311 nm (Figure 3).



Figure 3. Absorption spectrum of ferulic acid in the UV/Vis region

3.2.2 Selectivity

The spectra obtained by the FA solution between the length of 200 to 600 nm and for a solution composed only for Soluplus® present in the solid dispersion, were compiled and are represented in Figure 4.

Depending on the spectra obtained, it can be affirmed that the validation method is adequate in terms of selectivity, considering that there are no absorption peaks for Soluplus® at the same wavelength characteristic of FA, thus there is no interference of the polymer to the drug.



Figure 4. Spectrum in the UV region of ferulic acid and Soluplus®

3.2.3 Linearity

The analytical curve was constructed according to the resolution recommendations, with seven concentration points, in the content range of 80 to 120%. The means of absorbance values and RSD (%) for each concentration are in Table 2.

From these values, the mean analytical curve was obtained for the quantification of FA, the equation of the line and the value of the r found, as can be observed in Figure 5. Therefore, in line with the results of the analytical curve obtained in triplicate, there is an average correlation coefficient (r) value equal to 0.9994, which indicates a significant linear regression in accordance with RDC $n^{\circ}166$.

Concentration (µg/mL)	2	4	5	6	8	10	12
Absorbance	0.1550 0.1561 0.1611	0.3221 0.3124 0.3278	0.4008 0.3878 0.4036	0.4842 0.4742 0.5167	0.6418 0.6311 0.6226	0.8149 0.8185 0.7965	0.9907 0.9585 0.9723
Average	0.1574	0.3208	0.3974	0.4917	0.6318	0.8099	0.9738
RSD (%)	1.6865	1.9819	1.7322	3.6898	1.2432	1.1895	1.3544

Table 2. Absorbance values obtained in the construction of the analytical curve

of FA





3.2.4 Precision

The values for the mean of the actual concentration and the relative standard deviation (RSD) of this concentration for repeatability and intraday precision are arranged in tables 3 and 4, respectively. For the evaluation of this parameter, a maximum value of 5% of RSD was determined, and it was observed, therefore, that the method was accurate and was not influenced by the period that was performed or by the change of analyst.

Theoretical	Average Real	RSD (%)
Concentration (µg/mL)	Concentration (µg/mL)	
2	2.007	1.624
5	4.955	1.706
12	12.037	1.346

Analyst	Theoretical	Average Real	RSD (%)
	Concentration	Concentration	
	(µg/mL)	(µg/mL)	
Analyst 1	2	1.980	1.228
	5	4.975	1.734
	12	12.243	1.045
Analyst 2	2	2.135	1.686
	5	5.057	1.938
	12	12.118	1.407

Table 4. Values obtained for intra-day accuracy

3.2.5 Accuracy

Through the analysis of accuracy, it is possible to prove the proximity of the results of the method, with regard to the result admitted as true, that is, according to the actual concentrations. For this analysis, three different concentrations were admitted, high, medium and low, determining the concentration of each one. Thus, it was obtained values of accuracies between 99.11 and 100.36% (Table 5) and these in agreement with the true values and within the acceptable range that is 95-105% as established as a criterion for this parameter.

Theoretical	Average Real	Accuracy (%)	
Concentration (µg/mL)	Concentration (µg/mL)		
2	2.007	100.368	
5	4.955	99.115	
12	12.037	100.310	

Table 5. Accuracy values at three levels of FA concentration.

3.2.6 Detection and Quantification Limits

With the data obtained by the calibration curves, the limits of detection and quantification were calculated using equations 5 and 6, respectively. The resulting values were 0.4112 μ g/mL for DL and 1.2461 μ g/mL for QL.

3.3 Quantification of solid dispersions

The ferulic acid content in solid dispersions, yield and incorporation efficiency by the applied method are shown in Table 6. From the values obtained, it can be denoted that the ferulic acid content was satisfactory through the two techniques performed, as well as the incorporation efficiency had similar values. However, there was a highlight in the performance for the kneading technique, which allowed a considerable value if considered with the evaporation rotary.

Solid	Ferulic acid	Standard	Yield (%)	IE (%)
dispersion	content	deviation		
KND	4.28 mg/10mg	0.005	90.96%	85.6
	of SD			
ER	4.13 mg/10 mg	0.018	63.89%	82.6
	of SD			

Table 6. FA content in solid dispersions and efficiency of incorporation (%)

3.4 Chemical physical characterization

3.4.1 X-ray diffraction (XRD)

X-ray diffraction is one of the techniques widely used for the development and analysis of pharmaceutical products, being possible from this the measurement of the crystallinity of a material, through the graph which presents the relationship between the intensity of crystalline reflections and the diffraction angle used. Therefore, crystalline materials present well-defined and intense reflections, while those that do not show any or low intensity are considered as amorphous substances²³. In view of this importance, the behavior of isolated FA, the polymer used – Soluplus®, the physical mixture KND and ER method.

Based on the diffractogram obtained (Figure 6), it can be seen that FA shows a behavior of crystalline substance, with crystalline reflections of high intensity at 12.7°, 15.4°, 26.4°, 29.3°, 44.10° and 64.3°, as well as several others of lower intensity at 10.5°, 17.2°, 24.5°, 31.3°, 35.9°, 39°, similar results were demonstrated in the study by Wang et al¹⁰. This crystallinity presented by FA corroborates the low aqueous solubility of the drug. With regard to Soluplus® copolymer the presence of a halo pattern, which is characteristic of amorphous powders, is denoted²⁴.

Although the crystalline reflections of FA have been slightly reduced in PM with Soluplus®, it is also observed that there is the presence of intense peaks characteristic of the drug, as in 12.8°, 15.4° and 26.4°. Unlike what occurs in SDs by KND and ER that there is a considerable reduction in the intensity of these peaks, as well as in the peaks at 44.10° and 64.3°, in addition, the disappearance of peaks in 17.2°, 21.1° and 24.5° which are lower. Thus, it is understood that the methods KND and ER applied for the development of SD promoted a greater process of amorphization of the drug and that there were no significant differences between the two systems obtained.

Unlike the result obtained by Moreira et al²⁵, in a development of SDs with Hecogenin and five polymers, this drug did not present a good interaction with Soluplus® which was obtained a material with similar reflections of the compound and without reducing the intensity considerably, presenting itself as the best polymer for this development - the HPMC, in contrast the present study demonstrates that Soluplus® was able to form solid dispersion with ferulic acid of the methods applied.



Figure 6. FA diffractograms, Soluplus®, PM, KND and ER methods.

3.4.2 Fourier transform infrared spectroscopy (FTIR)

Infrared spectroscopy is correlated with the absorption of electromagnetic energy by molecules and depending on the different chemical structures and concentration of each, these will confer different energy absorptions²⁶. Using this method assumes that the mixing of two components at the molecular level should lead to changes, this are manifested in the spectrum in the shape changes, in the frequency and width of the bands through the interaction between drug and polymer²⁷.

Figure 7 shows the FTIR spectrum of FA, Soluplus®, physical mixture and solid dispersions obtained by KND and ER techniques. With regard to FA, a band in 3434 cm⁻¹ corresponding to axial deformation of the hydroxyl group (O-H) and another band in 3016 cm^{-1} attributed to the asymmetric stretching of the C-H group can be observed in the most energetic region of the spectrum. In 1690 cm⁻¹, the characteristic band was observed to carbonyl and three intense bands were also exhibited in 1617,1510 and 1427 cm⁻¹ corresponding to axial deformation C=C of the aromatic ring of FA. In addition,

bands were verified in 1269 cm⁻¹ and 1205 cm⁻¹, respectively, corresponding to the asymmetric stretch C-O-C and stretch C-OH^{18,28}.



Figure 7. FTIR Spectra for FA, Soluplus®, PM, KND and ER methods

The spectra obtained for the physical mixture and the SDs developed, it can be denoted that the spectral profile of ferulic acid was maintained and Soluplus®, without the appearance of new bands, however, there are reductions in the intensity of certain bands, mainly in solid dispersions such as, in the 1730 cm⁻¹ region which represents the carbonyl strain band of Soluplus®, in consideration of the 1617 cm⁻¹ band present in the FA molecule by the vibration of the stretch in the aromatic nucleus, as well as in 1427 cm⁻¹ and in the bands at 804 and 800 cm⁻¹ there is a considerable reduction in the intensity of these peaks, especially in the SDs, which are characteristic of hydrogen atom adjacent to the phenyl group in the FA molecule, can infer that there is a greater interaction through hydrogen bonds in this region. In addition, it is important to highlight that there is enlargement and more intense reduction of the band in the region between 3000 and 2800 cm⁻¹ in the spectra of the systems obtained by solid dispersion methods (KND and ER) and that in this region there is the band of 2930 cm⁻¹ corresponding to the aromatic C-H

bond in Soluplus[®], and it can be inferred that in this group there are significant interactions with FA and being more evident in the SD by ER.

From this analysis it can be understood that there is greater interaction of FA with the polymer matrix when the methods of obtaining solid dispersions by KND and ER methods are applied, considering that the reduction of band intensity was more visible when compared to the physical mixture.

3.4.3. Scanning electron microscopy (SEM)

Analyses of the photographs of pure ferulic acid, physical mixture and solid dispersions obtained by ER and KND, Figure 8A demonstrates the crystalline behavior of PA, as needle-like crystals, corroborating the result exposed by Wang et al¹⁰. This morphology is perceived by substances that present low aqueous solubility, due to its well organized and symmetrical structure of molecules which thus hinder the penetration of water. With regard to Soluplus® particles, according to the literature, morphology is presented with cluster-free aspects and is characterized more as an amorphous substance, besides presenting a rough aspect^{29,25}.

PM's photomicrograph (Figure 8B) demonstrates that it is already possible to obtain a good interaction between the drug and polymer through this technique, not being visualized the crystalline profile of the pure drug, this same characteristic is observed in the micrographs of the SDs obtained by the methods of KND and ER, in addition, is presented the formation of a polymer mesh in which it can be inferred that the FA is dispersed , thus demonstrating a good interaction between the two components for solid dispersion formation. This result corroborates those expressed by the XRD, in which through the physical mixture the reduction of crystalline reflexes was obtained, however, this reduction was more expressive for the solid dispersions systems developed by the methods of KND and ER, therefore, involving a greater process of amorphization of the drug that will corroborate with an increase in its solubility.



Figure 8. SEM of FA (A), PM (B), KND (C) and ER (D) with increase in 500x

3.4.4. Differential Exploratory Calorimetry

According to the calorimetric curve of FA (Figure 9), it is noted the presence of two important events, the first being related to the characteristic fusion event of crystalline substance, having as melting temperature (Tpeak) at 173.4 °C, with Tonset at 171.6 °C and Tendset at 175,84 °C and $\Delta H = -132.53$ J/g, this result being consistent with that described in the literature by the Japanese Pharmacopoeia³⁰, which accepts melting temperatures for this drug between 173 and 176°C. In addition, the second endothermic event observed is related to the degradation of FA mainly related to the oxidation process, which has as Tonset 184,.9 °C and Tendset 233.,12°C, with peak temperature at 224.,78 °C. Similar results are presented by Bezerra et al³¹.

Soluplus® presents a single endothermic event in the temperature range used, with Tonset at 47.,93 °C, Tendset at 109.,32 °C and Tpeak at 73.,5°C, which is related to copolymer dehydration, expressing equivalent result to the study developed by Prakash and Pravin³².

With regard to the calorimetric curves for PM and SDs, an endothermic event with variation of enthalpy is observed indicating the possible fusion of the asset, however, in a subtle way due to the amorphization process resulting from the methodologies applied and interactions between the drug and polymer, similar findings are observed in the results exposed by the diffuser (Figure 9) and in the study developed by Zhang et al³³, in a study SD of Simvastatin and Soluplus[®]. For the physical mixture it is said that Tpeak occurs at 143 °C with enthalpy variation of -37.94 J/g; sd by KND method the Tpeak is at 158.53 °C and $\Delta H = -26.48$ J/g and for Tpeak at 160.81 °C and $\Delta H = -11.97$ J/g, it is important to highlight that for the solid dispersion obtained by the ER method there is a variation of smaller enthalpy and it can be inferred that the solubilization of the drug in the polymer in this system occurred in a smaller way, understanding that there was a more representative interaction between the two components, corroborating this result with that obtained by the FTIR spectrum, which presents bands with lower intensities for this particular acquisition.

In view of this approach, it can be affirmed that the KND and ER techniques allowed a process of change from the crystalline state to the amorphous of FA, and it is not possible to observe the peaks of fusion of the asset in the systems obtained. This amorphation process corroborates the results of the XRD, as well as with studies developed by Alves et al³⁴ that developed solid dispersions by the same methodologies using PVP K30 and efavirenz, as well as the work developed by Yadav et al³⁵.



Figure 9. Calorimetric curves of FA, Soluplus®, PM, KND and ER methods

4. Conclusion

In the present study, the solubility diagram indicated that Soluplus[®] provided the best increase in the solubility of ferulic acid, demonstrating to be the best among all polymers tested. The validation of the methodology obtained was selective for quantification of FA, with proven precision and accuracy. In addition, the kneading technique and the evaporation rotary were effective, obtaining a satisfactory content FA of SD. The decrease in the crystallinity of the drug was confirmed through the chemical physical characterization, mainly by the results of the XRD and DSC, indicating the performance of solid amorphous dispersion, where the ER method presented a more expressive result, based on the FTIR and DSC, and this fact can be understood because for this method a greater amount of solvent is used to obtain, in which it can promote a greater and better interaction between the components producing , in this way, a more amorphous solid dispersion.

As future perspectives, additional research needs to be carried out, such as robustness, thermogravimetric analysis and the in vitro dissolution study. In moreover, the of in vitro antioxidant activity of solid dispersions.

5. References

- Fridgeirsdottir GA, Harris R, Fischer PM, Roberts CJ. Support Tools in Formulation Development for Poorly Soluble Drugs. *J Pharm Sci.* 2016;105(8):2260-2269. doi:10.1016/j.xphs.2016.05.024
- 2. Wu CY, Benet LZ. Predicting drug disposition via application of BCS: Transport/absorption/ elimination interplay and development of a biopharmaceutics drug disposition classification system. Pharm Res. 2005;22(1):11-23. doi:10.1007/s11095-004-9004-4
- Kambayashi A, Kiyota T, Fujiwara M, Dressman JB. PBPK modeling coupled with biorelevant dissolution to forecast the oral performance of amorphous solid dispersion formulations. *Eur J Pharm Sci.* 2019;135(March):83-90. doi:10.1016/j.ejps.2019.05.013
- Di L, Fish P V., Mano T. Bridging solubility between drug discovery and development. *Drug Discov Today*. 2012;17(9-10):486-495. doi:10.1016/j.drudis.2011.11.007
- Williams HD, Trevaskis NL, Charman SA, et al. Strategies to address low drug solubility in discWILLIAMS, Hywel D.; TREVASKIS, Natalie L.; CHARMAN, Susan A.; et al. Strategies to address low drug solubility in discovery and development. Pharmacological Reviews, v. 65, n. 1, p. 315–499, 2013.overy and . *Pharmacol Rev.* 2013;65(1):315-499. doi:10.1124/pr.112.005660
- Tran P, Pyo YC, Kim DH, Lee SE, Kim JK, Park JS. Overview of the manufacturing methods of solid dispersion technology for improving the solubility of poorly water-soluble drugs and application to anticancer drugs. *Pharmaceutics*. 2019;11(3):1-26. doi:10.3390/pharmaceutics11030132
- Vo CLN, Park C, Lee BJ. Current trends and future perspectives of solid dispersions containing poorly water-soluble drugs. *Eur J Pharm Biopharm*. 2013;85(3 PART B):799-813. doi:10.1016/j.ejpb.2013.09.007
- 8. Knopp MM, Nguyen JH, Becker C, et al. Influence of polymer molecular weight on in vitro dissolution behavior and in vivo performance of celecoxib:PVP

amorphous solid dispersions. *Eur J Pharm Biopharm*. 2016;101:145-151. doi:10.1016/j.ejpb.2016.02.007

- Zduńska K, Dana A, Kolodziejczak A, Rotsztejn H. Antioxidant properties of ferulic acid and its possible application. *Skin Pharmacol Physiol*. 2018;31(6):332-336. doi:10.1159/000491755
- Wang J, Cao Y, Sun B, Wang C. Characterisation of inclusion complex of transferulic acid and hydroxypropyl-β-cyclodextrin. *Food Chem.* 2011;124(3):1069-1075. doi:10.1016/j.foodchem.2010.07.080
- 11. Mancuso C, Santangelo R. Ferulic acid: Pharmacological and toxicological aspects. *Food Chem Toxicol*. 2014;65:185-195. doi:10.1016/j.fct.2013.12.024
- Ghosh S, Basak P, Dutta S, Chowdhury S, Sil PC. New insights into the ameliorative effects of ferulic acid in pathophysiological conditions. *Food Chem Toxicol.* 2017;103:41-55. doi:10.1016/j.fct.2017.02.028
- de Paiva LB, Goldbeck R, dos Santos WD, Squina FM. Ferulic acid and derivatives: Molecules with potential application in the pharmaceutical field. *Brazilian J Pharm Sci.* 2013;49(3):395-411. doi:10.1590/S1984-82502013000300002
- 14. Saija A, Tomaino A, Trombetta D, et al. In vitro and in vivo evaluation of caffeic and ferulic acids as topical photoprotective agents. *Int J Pharm.* 2000;199(1):39-47. doi:10.1016/S0378-5173(00)00358-6
- Martins CR, Lopes WA, De Andrade JB. Solubilidade das substâncias orgânicas. *Quim Nova*. 2013;36(8):1248-1255. doi:10.1590/S0100-40422013000800026
- Chaves Júnior JV, dos Santos JAB, Lins TB, et al. A New Ferulic Acid– Nicotinamide Cocrystal With Improved Solubility and Dissolution Performance. J Pharm Sci. 2019. doi:10.1016/j.xphs.2019.12.002
- 17. Jéssica Cabral Ferreira. DESENVOLVIMENTO E VALIDAÇÃO DE UM
 MÉTODO POR CLUE-DAD PARA IDENTIFICAÇÃO E QUANTIFICAÇÃO
 DO ÁCIDO FERÚLICO E SEUS PRODUTOS DEGRADAÇÃO FORÇADA. J

Chem Inf Model. 2013;53(9):1689-1699. doi:10.1017/CBO9781107415324.004

- Nadal JM, Gomes MLS, Borsato DM, et al. Spray-dried solid dispersions containing ferulic acid: comparative analysis of three carriers, in vitro dissolution, antioxidant potential and in vivo anti-platelet effect. *Drug Dev Ind Pharm*. 2016;42(11):1813-1824. doi:10.3109/03639045.2016.1173055
- De DEJ. Ministério da Saúde MS Agência Nacional de Vigilância Sanitária -ANVISA. 2017;2017.
- Shamma RN, Basha M. Soluplus®: A novel polymeric solubilizer for optimization of Carvedilol solid dispersions: Formulation design and effect of method of preparation. *Powder Technol.* 2013;237:406-414. doi:10.1016/j.powtec.2012.12.038
- Reintjes T. 10. KolliphorTM P grades (Poloxamers). Solubility Enhanc with BASF Pharma Polym Solubilizer Compend. 2011:103-111.
- Psimadas D, Georgoulias P, Valotassiou V, Loudos G. Molecular Nanomedicine Towards Cancer : *J Pharm Sci.* 2012;101(7):2271-2280. doi:10.1002/jps
- Chadha R, Bhandari S. Drug-excipient compatibility screening-Role of thermoanalytical and spectroscopic techniques. *J Pharm Biomed Anal.* 2014;87:82-97. doi:10.1016/j.jpba.2013.06.016
- 24. Papadimitriou SA, Barmpalexis P, Karavas E, Bikiaris DN. Optimizing the ability of PVP/PEG mixtures to be used as appropriate carriers for the preparation of drug solid dispersions by melt mixing technique using artificial neural networks: I. *Eur J Pharm Biopharm*. 2012;82(1):175-186. doi:10.1016/j.ejpb.2012.06.003
- 25. de França Almeida Moreira CDL, de Oliveira Pinheiro JG, da Silva-Júnior WF, et al. Amorphous solid dispersions of hecogenin acetate using different polymers for enhancement of solubility and improvement of anti-hyperalgesic effect in neuropathic pain model in mice. *Biomed Pharmacother*. 2018;97(August 2017):870-879. doi:10.1016/j.biopha.2017.10.161
- 26. RAPOSO DE MELLO D. Farmacopeia Brasileira. Farm Bras 5^a edição. 2010;2:1-

523. www.anvisa.gov.br.

- Neves de Lima AA, Souto dos Santos PB, Marques de Lyra MA, Araujo dos Santos FL, Rolim-Neto PJ. Solid dispersion systems for increase solubility: cases with hydrophilic polymers in poorly water soluble drugs. *Brazilian J Pharm*. 2011;92(4):269-278.
- Mathew S, Abraham TE. Physico-chemical characterization of starch ferulates of different degrees of substitution. *Food Chem.* 2007;105(2):579-589. doi:10.1016/j.foodchem.2007.04.032
- 29. Tousif Ayyub K, Moravkar K, Maniruzzaman M, Amin P. Effect of melt extrudability and melt binding efficiency of polyvinyl caprolactam polyvinyl acetate polyethylene glycol graft copolymer (Soluplus®) on release pattern of hydrophilic and high dose drugs. *Mater Sci Eng C*. 2019;99(January 2018):563-574. doi:10.1016/j.msec.2019.01.126
- 30. PMDA. The Japanese Pharmacopoeia 16th Ed. 2011;2014(47):2319. http://www.pmda.go.jp/english/pharmacopoeia/pdf/jpdata/JP16eng.pdf.
- Bezerra GSN, Pereira MAV, Ostrosky EA, et al. Compatibility study between ferulic acid and excipients used in cosmetic formulations by TG/DTG, DSC and FTIR. J Therm Anal Calorim. 2017;127(2):1683-1691. doi:10.1007/s10973-016-5654-9
- Kendre PN, Chaudhari PD. Effect of amphiphilic graft co-polymer-carrier on physical stability of bosentan nanocomposite: Assessment of solubility, dissolution and bioavailability. *Eur J Pharm Biopharm*. 2018;126:177-186. doi:10.1016/j.ejpb.2017.06.024
- Zhang Y, Liu Y, Luo Y, et al. Extruded Soluplus/SIM as an oral delivery system: characterization, interactions, in vitro and in vivo evaluations. *Drug Deliv*. 2016;23(6):1902-1911. doi:10.3109/10717544.2014.960982
- 34. Alves LDS, De La Roca Soares MF, De Albuquerque CT, et al. Solid dispersion of efavirenz in PVP K-30 by conventional solvent and kneading methods. *Carbohydr Polym.* 2014;104(1):166-174. doi:10.1016/j.carbpol.2014.01.027

35. Yadav PS, Kumar V, Singh UP, Bhat HR, Mazumder B. Physicochemical characterization and in vitro dissolution studies of solid dispersions of ketoprofen with PVP K30 and d-mannitol. *Saudi Pharm J.* 2013;21(1):77-84. doi:10.1016/j.jsps.2011.12.007