

THIOMERS; A BLESSING TO EVOLUTING ERA OF PHARMACEUTICALS

Muhammad Zaman*^{1,2}, Muhammad Hanif¹, Sundas Qureshi²,

¹ Department of Pharmacy, Bahauddin Zakariya University, Multan, Pakistan

² Faculty of Pharmacy, The University of Lahore

Corresponding Author

Muhammad Zaman

Department of Pharmacy

Bahauddin Zakariya University, Multan, Pakistan

Email Address: m.zaman2157@gmail.com

Contact: 0092-3006095928

Abstract

Thiomers are the polymers modified for the mucoadhesive properties and other additive properties by incorporating thiol moieties in the backbone of the unmodified polymeric chain by exchange reactions or simple oxidation reactions. Drug that are less soluble and permeable can be complexed with thiomers for their increased absorption through the mucosal membranes by increase in contact time and prolonged stay in body due to mucoadhesion. Immobilization of thiol group therefore increases the mucoadhesive properties of the modified polymer by 2-140 folds. The prepared thiomers are characterized and made stable by different techniques. Thiomers also give the controlled delivery of the active in the body. Different polymers that are modified by thiolation are chitosan, polyacrylic acid, sodium alginate, sodium carboxy methyl cellulose, guar gum etc. Thiomic formulations are a challenge to deliver drugs with low therapeutic compatibility. Micro- and nano- preparations containing thiomers can be prepared by different techniques such as covalent crosslinking, insitu gelation, radical emulsion polymerization, emulsification etc. Nowadays thiomers have wide range of applications as a promising pharmaceutical excipient in the evaluating era of pharmaceutical technology.

Keywords: Thiomers, Thiolation, Mucoadhesion

Introduction

Bioadhesion is the phenomenon by which any molecule adheres to the biological membrane but when the adherence to the mucus is specified it is referred to as mucoadhesion[1]. Mucoadhesion is introduced in the pharmaceutical field for more than 40 years and it is of main importance in oral delivery systems, as drugs taken from oral route have various bioavailability issues[2, 3]. Adhesion to the membranes is of importance as it prolongs the stay of the drug in the body subsequently increasing bioavailability of the drug which in turn improves patient compliance[4]. The natural or synthetic macromolecules are present that adhere to the mucus layer by interacting with the glycoproteins in the mucus, not by covalent bonding, but by weak ionic interactions such as, hydrogen bonding and Vander Waal's forces[1, 5]. Nowadays, efforts are being made to increase the mucoadhesive properties of the polymer such as improved resistance to enzymes, increased cohesive force and facilitated paracellular diffusion of the drug, increased bioavailability and increased patient compliance[2, 6]. For this purpose, the new generation of mucoadhesive polymers developed by integrating sulfhydryl groups on the backbone of the polymer resulted in thiolated polymers, also called as thiomers[6, 7]. This structural modification in the polymer has led to the improvement in the mucoadhesive properties of the polymers by 2- to 140- folds[8]. Polymers modified are

- Alginate
- Chitosan [2]
- Polycarbophil[9]
- Polyacrylic acid [0]
- Xyloglucan [1]
- Carboxy methyl cellulose[12]
- polyaspartamide[13, 14]
- hydroxyethyl cellulose[15]

Types of Thiomers

Followings are the types of thiomers

- **Cationic thiomers**
- **Anionic thiomers**

Table 1: Examples of cationic and anionic polymers

Cationic thiomers	Anionic thiomers
Hydroxyethyl cellulose	Poly acrylic acid cystamine conjugate
	Sodium carboxy methyl starch (CMS)
	Carboxy methyl guar gum (CMG)
	Carboxymethyl cellulose
	Sodium alginate
Chitosan	

Cationic thiomers

Basically these are the thiomers based on chitosan, prepared by immobilizing thiol group on 2-amino position of the glucosamine, present in the polymer chain. Examples include chitosan-cysteine, chitosan-thioglycolic acid, chitosan-thiobutylamidineetc (table 1) [2].

Chitosan

Chitosan, a natural mucoadhesive polysaccharide polymer, obtained from deacetylation of chitin present in the crustacean shell, is chemically named as [poly {(1, 4)-2-amino-2-deoxy-d-glucopyranose}]. It consists of two subunits N-acetylglucosamine and glucosamine[16]. Chitosan has the mucoadhesive and permeation enhancing properties due to interaction of the amino group with the anionic structures in the mucus or epithelial layer due to its polycationic nature[17]. The primary amino group present in glucosamine is easily protonated in acidic media which increases its solubility. While it remains stable in basic and neutral media[2, 18].

Modified generation of chitosan is prepared with more pronounced mucoadhesive properties by incorporating thiol group on the backbone of the polymer that is creating the thiolated chitosan[19]. Thiolated chitosan are with improved mucoadhesive properties, enhanced permeation and in situ gelling properties such as chitosan thio butyl amidine conjugate, chitosan cysteine conjugate, chitosan thioglycolic acid conjugate etc.[2, 20].

Increased mucoadhesive property of chitosan

Thiolated chitosan is the modified polymer having ability to bind with the mucin through covalent bonding rather than weak ionic interactions such as hydrogen bonding or vanderwaal's forces as simple chitosan. Sulphur present in the thiolated chitosan make disulfide bonding with the cysteine rich domain of the mucin thus increasing time to stay in body which in turn

increases bioavailability and also may help to reduce dosing frequency which ultimately increase patient compliance[2].

Mucin

Mucin is a glycoprotein consisting of protein core with branched oligosaccharides over 63% of its length, makes 5% of total mucus. It accounts for gel like properties of the mucus. Cysteine present in the mucin glycoprotein account for 1.5% of the amino acid in the small intestine .i.e. about 9 $\mu\text{mol SH/g}$ of mucus[2].

Preparation of thiolated chitosan

Thiolated chitosan is prepared by introducing the sulfhydryl bearing moiety on the 2-amino position of the glucosamine subunit. This is done by reacting carboxylic group of the moiety such as cysteine or thioglycolic acid with the amino group in the polymer back bone leading to formation of the amide bond or amidine bond, in the presence of 1-ethyl-3-(3-dimethylaminopropyl) carbodiimides (EDAC) or thiobutyric acid, respectively, as coupling agent. Thiol group may undergo oxidation so synthesis must be done under inert conditions or at $\text{pH} < 5$, as at this pH the concentration of the thiol anions is low so disulfide formation seldom occurs[2, 4, 21].

The schemes of reactions involved in the formation of thiolated chitosan are shown in the figure below. It shows two schemes indicating one step process and two steps process. Single step processing of chitosan to achieve thiolated chitosan shows reaction medium consisting of water. The thiolation takes place in the presence of carbodiimide (EDAC) and NHS (N-hydroxysuccinimide). While the second process involving the two steps; first step involves DMF (N,N-dimethylformamide) as a reaction medium to avoid hydrolysis of the ester formed thereby in the reaction. This scheme is shown in figure 1[18].

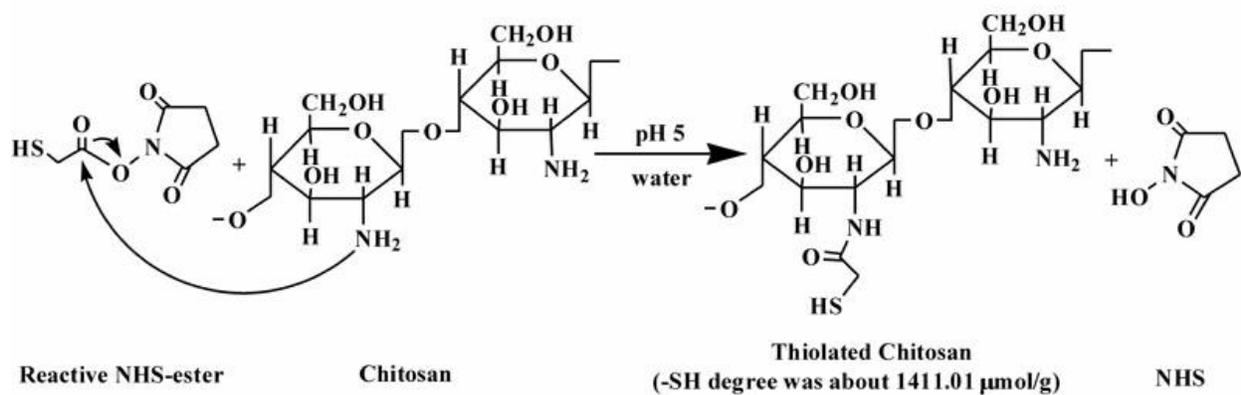
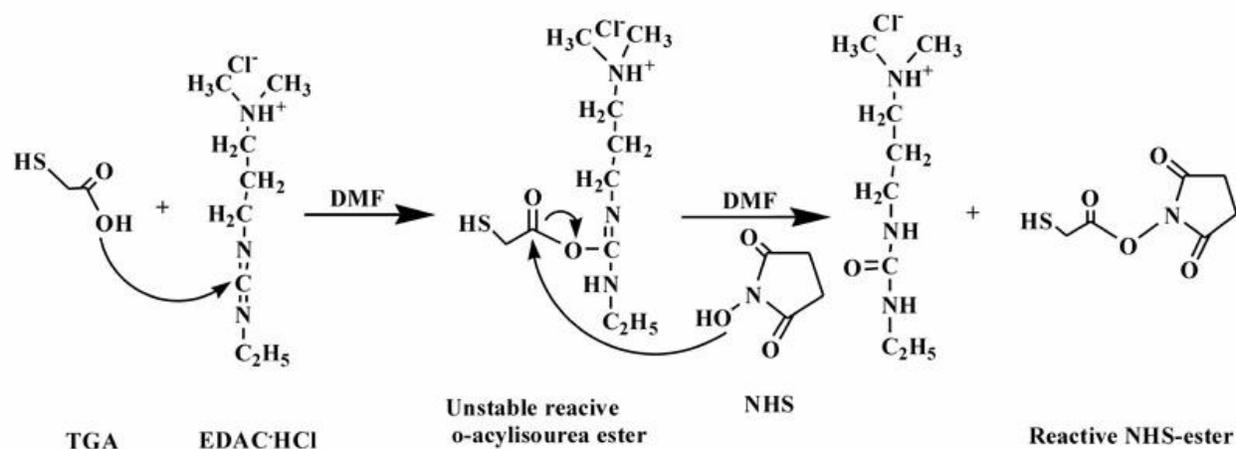
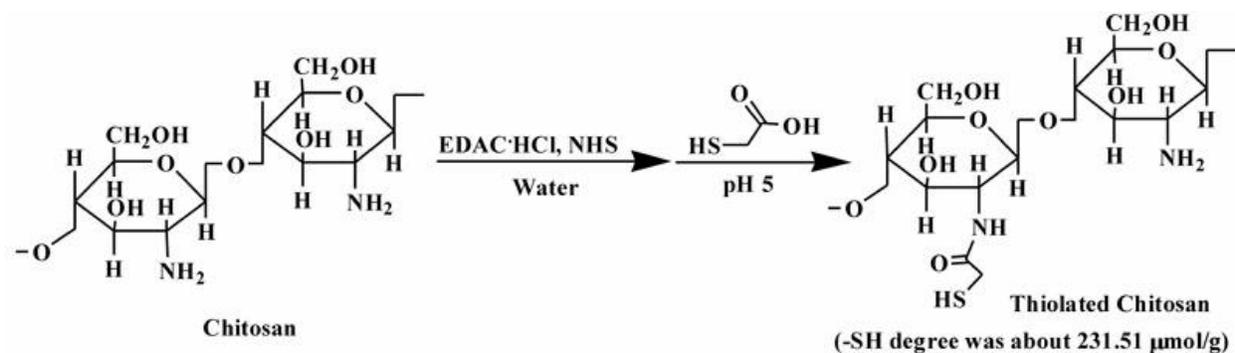


Figure 1: Process for Thiolation of Chitosan

Degree of effective thiol immobilization

Studies have shown that immobilization of 25- 250 μmol thiol groups per gram of the chitosan are effective in improving the mucoadhesive properties[22] and permeability of the chitosan polymer. If the thiol groups are reduced their immobilization can be determined by Ellman's

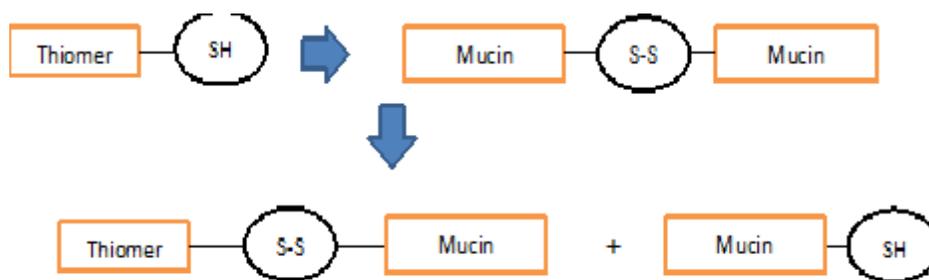
reagent, while if they are oxidized then by borohydride. Alternatively iodine titration method can be used[4].

Effect on mucoadhesion

Mucoadhesive properties of the chitosan are enhanced by 5-10 folds in case of thioglycolic acid conjugate and 10-20 fold in case of chitosan-4- thiobutylamide conjugate[2, 23]. The main perspective of thiolation is to increase the mucoadhesion. Mucoadhesion of different polymers increase by many fold with the addition of thiol group (Table 2) [24, 25].

Mechanism of mucoadhesion of thiomers[2]

Mucin is present all over the mucus membrane which has cysteine rich domains. The thiol group interact with these cysteines and lead to formation of the disulfide bonds either by oxidation of the thiol groups or by thiol/disulfide exchange reaction (Figure 2).



Mechanism of mucoadhesion of thiomers

Figure 2: Mechanism of Mucoadhesion of Thiomers

The extent to which the mucoadhesion takes place depends upon the pKa of the thiol group, pH of thiolated chitosan and pH of the surrounding medium. The disulfide bonds thus formed are not influenced by the ionic strength and pH condition.

In-situ gelling of chitosan[26]

Ability of chitosan to crosslink adds more to its mucoadhesive efficiency. The chitosan, after forming surface interaction, it starts to form bonds with itself leading to more strong adhesion. The insitu gelling property of chitosan has been of great value for nasal, vaginal and other preparations[2].

Thiolated chitosan has shown more pronounced gelling property in comparison to unmodified chitosan. The thiol groups interact with each other leading to formation of more strong gel. This occurs due to oxidation of the thiol group at physiological pH. At pH 5.5, the sol-gel transition of the thiolated chitosan occurs after 2h. When the thiolated chitosan is rheologically observed for the gelling property, it has shown significant decline in the thiol group concentration showing

formation of disulfide bond formation. Also, thiolated chitosan show more elasticity in comparison. The insitu gel forming property of thiolated chitosan is important in case of liquid or semisolid preparation for nasal, vaginal or ocular use, where thin layer of solution is applied which form gelly layer on mucus and thereby increase the contact time and thus bioavailability of the drug in the body [2].

Effect on permeation

Thiolation of chitosan enhances its permeation effect more folds as compared to the simple chitosan. It can be assayed by using Ussing's chamber, through the fresh intestinal mucosa. The studies have shown increase in the uptake of flouresenced bacitracin by 1.6 folds using chitosan cysteine, while for cysteine-TBA it has shown increase of cationic marker rhodamine 123 uptake by 3 folds as compared to unmodified chitosan[2].

Mechanism of permeation enhancement

Permeation enhancement effect of the thiolated chitosan can be explained by its effect on protein tyrosine phosphatase, the enzyme that is responsible for dephosphorylation of the transmembrane occludin tyrosine, the protein responsible for opening and closing of these tight junctions between cells. Thiolated chitosan inhibit this dephosphorylation to increase the permeation for drug, thus increasing bioavailability[2].

Thiolated chitosan in different dosage forms

Microparticles

Microparticulate dosage forms due to their small size have the ability to increase the bioavailability due to their increased time to stay in the body. Now when thiolated chitosan with improved mucoadhesive property and being more resistant to degradation is incorporated, the dosage form with more pronounced properties will be obtained[2]. Microparticles of thiolated polycarbophil were also prepared and shown to have 3 folds mucoadhesion as compared to unmodified polycarbophil[27].

Tablets

40% of all formulations are tablets. Thiolated chitosan can be incorporated for the prolonged release effect to the dosage form due to its insitu gelling properties. If only drug is mixed with polymer and compressed into tablet, it can still show zero order release for several hours. It shows hydration and diffusion account for the release rate[2].

Solutions

Thiolated chitosan seems to be unstable when in solution form due to its gelling property and disulphide bond formation within the polymer. This polymer used in solution formulations for eye can be effective in the most common eye problem, dry eye, the condition in which mucous

layer of eye acting as a surfactant becomes defective, there it can be used to form protective layer to cope this condition[2].

Gels

Thiolated chitosan gel formulation seems to be very effective as compared to simple chitosan gels due to the thiolated back bone which enhance its mucoadhesive properties[2].

Hydroxyethyl cellulose

Hydroxyethyl cellulose-cystamine is synthesized by open ring oxidative amination of the hydroxyethyl cellulose to incorporate thiol groups on the polymer chain. The modified polymer thus obtained has 3 fold more viscosity in comparison to the unmodified hydroxyethyl cellulose. Also the swelling properties and the mucoadhesion is markedly improved[18, 28].

Anionic thiomers

Carboxylic acid group is present as the substrate in anionic thiomers. Presence of carboxylic groups makes the immobilization of the thiol groups to these polymers more efficient by amide bond formation in the presence of carbodimides. Examples include poly acrylic acid-cysteamine, CMC-cysteine, alginate-cysteine (Table 1)[13].

Sodium alginate cysteine conjugate

Sodium alginate is thiolated for surplus effects. In the presence of carbodimidethiolation takes place (Figure 3) and the thiomers thus produced is studied and shown to have 50% more viscosity in comparison to the simple sodium alginate solution at 37⁰c. The swelling index is also improved [29]. Tablets thus prepared by using this modified polymer has shown to be more stable and resident on mucosa [11].

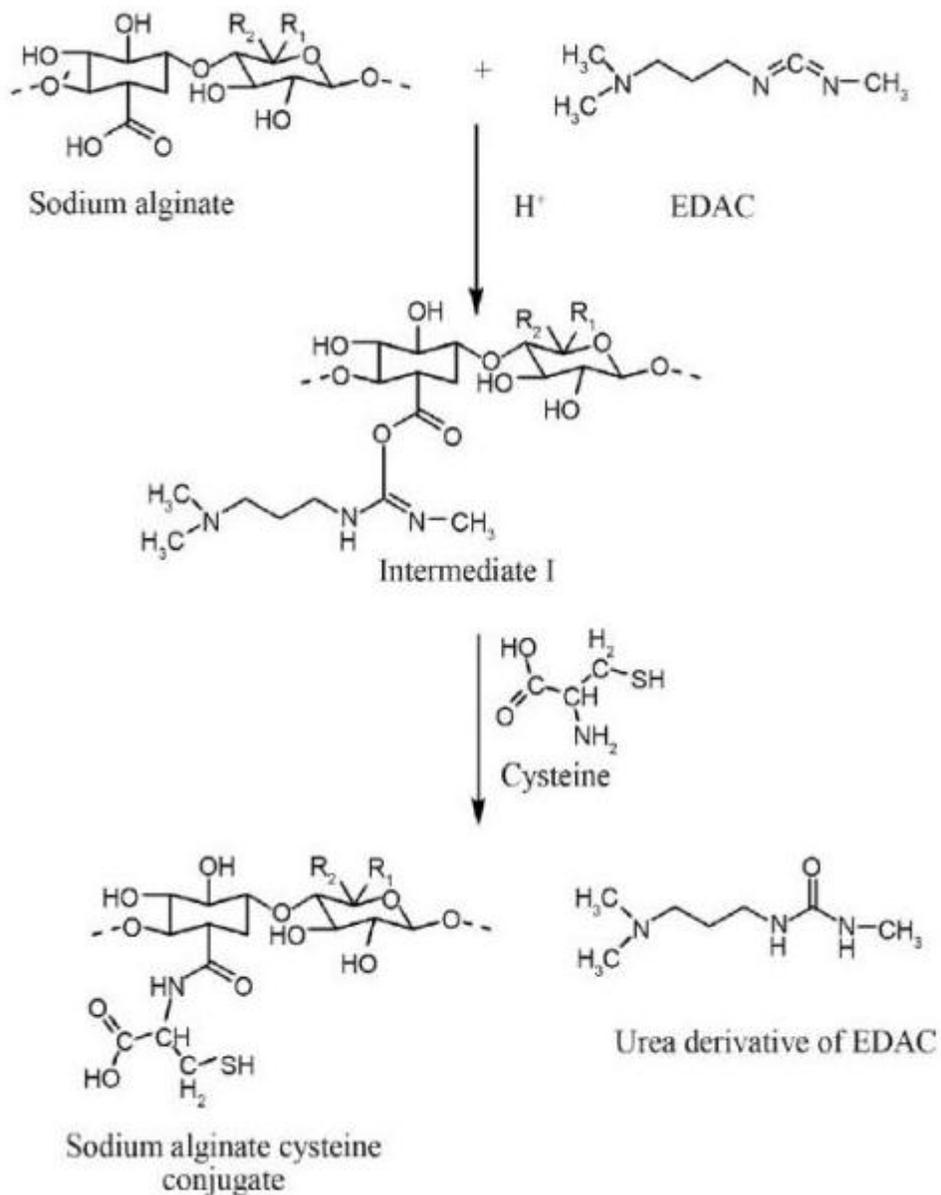


Figure 3: Process for Thiolation of Sodium Alginate

Poly acrylic acid cystamine conjugate

Poly acrylic acid cystamine conjugates are prepared in the presence of carbodimide by hydrolyzing polyacrylic acid polymer in demineralized water. Hydrolyzed polymer shows up its carboxylic acid group for S-S conjugation [30].

Polyacrylic acid homocysteine conjugates prepared in the presence of carbodimide have shown to contain sulfur $930 \mu\text{mol} \pm 83$ per gram of thioimer. Polyacrylic acid homocysteine has shown to have improved mucoadhesive characteristics as compared to unmodified polymer.

Sodium carboxy methyl starch (CMS) and carboxy methyl guar gum (CMG)[38]

Their thiomers are prepared in the presence of carbodimide by coupling reaction. Thiol groups in the form of cysteine are conjugated. Thereafter the mucoadhesive properties of both the polymers are studied which showed CMS to be less mucoadhesive than CMG. On the other hand the swellability of CMG is not affected by cysteine immobilisation. On the whole CMG proven to be a better candidate as release of drug from the matrix tablet is 1.5 fold than CMS.

Thiolated xyloglucan

Xyloglucans are important glycans that are cellulose microfibrils obtained from the seeds of the dicots. It is thiolated and its thiolation can be determined by Ellman's reagent while mucoadhesive extent can be determined by in-situ gelling of the modified polymer[5].

Table 2: Thiomers with their effect on mucoadhesion[13]

Thiomer	Increase in mucoadhesive strength
Chitosan iminothiolane	250 times
Poly acrylic acid-cysteine	100 times
Poly acrylic acid-homocysteine	Approx. 20 times [43]
Chitosan thioglycolic acid	10 times
Chitosan thioethylamidine	9 times
Alginate cysteine	4 times
Polymethacrylic acid-cysteine	Improved cohesiveness and mucoadhesion [44, 45]
Sodium carboxy methyl cellulose-cysteine	Increased mucoadhesion
Gellan gum-2-iminothiolane	Increased mucoadhesion[46]
Carbopol 980	Increased mucoadhesion [47]
Hyaluronic acid-L-cysteine	Increased mucoadhesion [48]
Carboxymethyl-Hyaluronic-Acid	Improved cohesiveness and mucoadhesion [49]
poly(ethyleneglycol)-glutathione	Improved drug delivery[50, 51]
Sodium alginate	Improved drug delivery [52]

Properties of thiomers

Hydrophilic properties of thiomers due to the thiol group make the polyacrylic acid and chitosan more mucoadhesive[13]. Other properties include cohesive properties, mucoadhesive properties, enzyme inhibition and permeation enhancement[7].

Characterization of thiomers[30]

Thiomers are tested and characterized for their stability and effectiveness by various methods such as

1. Determination of thiol group content[14]

The thiol group content on the polymer was determined by iodometry according to given procedure. The amount of thiol group tells the degree of thiolation. Hydrating with demineralized water thiomers were taken in the iodine flask. The pH of the solution was adjusted by adding 1 M HCl and then standard solution of 0.1N iodine was added and shook for 30 minutes. Excessive iodine was titrated with 0.1 N sodium thiosulphate solution. Starch is used as an indicator.

Same procedure mentioned above is used for estimation of blank without thiomers. The degree of thiolation is determined by following formula.

$$\% \text{ Thiol group content} = \frac{[(\text{Blank}-\text{Proper}) \times 0.1 \times 0.066 \times 100]}{0.1 \times \text{weight of thiomers}}$$

2. Di-Sulfide bond formation[14]

200 mg of thiomers is hydrated with iodine in a flask and pH was adjusted between 2-3 using 1M HCl. Then 0.6 ml of 3% solution of sodium borohydride was added to the polymer and shaken for 15 min to hydrate all the disulfide bonds to free thiol groups. After this the mixture is neutralized by addition of 0.5 ml of 1M HCl. The thiomers before reduction and after reduction of thiol groups is subtracted to estimate disulfide content[31].

$$\% \text{ Disulfide group content} = \frac{[(B1-B2) \times 0.1 \times 0.066 \times 100]}{0.1 \times \text{weight of thiomers}}$$

3. Swelling behavior[14]

Thiomers prepared at different pH are compared. 30mg of thiomers was compressed into a disc of 5 mm diameter and dipped in demineralized water at 20^oc by placing at permeable bottom tubes.

4. Viscosity[14]

The viscometer is used to measure the viscosity of thiomers. 2% solution is prepared and temperature conditions of 25±1 were used at 20 rpm.

5. Extent of mucoadhesion- in-vivo

The freshly excised intestinal mucosa of the sheep was taken to study the extent to which thiomers bind to the mucosa. The mucosa was clamped upside down and the exposed area was about 0.785cm². Equal amounts of all formulations are used to equilibrate each part. Every sample mucosa was exposed for 2 minutes with formulation to ensure the intimate contact between the two. The weights are used to detach the mucosa clamped in the pan to another, the weights thus used to detach mucosa is expressed as detachment stress which will be used to estimate in vivo mucoadhesion of thiomers in dynes/cm².

$$\text{detachment stress} \left(\frac{\text{dynes}}{\text{cm}^2} \right) = m * \frac{g}{A}$$

Where

m=mass

g=acceleration due to gravity

A=area of tissue exposed

ROUTES OF DRUG ADMINISTRATION USING THIOMERS AS A CARRIER OF THE DRUGS

There are various routes of drug administration that can be used to deliver the drug using thiomers as carrier. They have various applications using these routes of administration. The thiomers preparations are available for oral, nasal, ocular, buccal delivery of the drug. In literature, it is studied that thiomers can be used to deliver the drug through transmucosal route, gastrointestinal route, buccal route, oral route, nasal route, ophthalmic route and vaginal route of drug delivery. Different routes of drug administration along with the drugs that can be used through these routes are discussed as follows;

Oral delivery of drug

Oral route of drug administration is considered as the safe and effective for various drugs. Effects of various drugs can be enhanced when they are administered through oral route using thiolated polymers such as increased stay of drug in intestine and prolonged contact, enhanced permeation, enzyme inhibition [32]. Bioavailability of poorly bioavailable drugs like calcitonin, insulin and low molecular weight heparin can be increased using thiomers based formulations. There are various application of thiolated polymers in enhancement of bioavailability of poorly bioavailable drugs through oral route for examples;

Calcitonin; Calcitonin bioavailability is improved by conjugating it with the chitosan in modified form as compared to simple one due to the enzyme inhibiting and permeation enhancement effect[33]. It led to 5% more decrease in the calcium level[9].

Insulin; bioavailability and effect of insulin can be enhanced using thiolated polymer as the carrier in the form of matrix tablets. Thiolated Polycarbophil using cysteine was used to carry the insulin in the form of tablets. This formulation showed 36% more decrease in the blood sugar level comparatively [9].

Low molecular weight heparin; Low molecular weight heparin is administered subcutaneously which is painful and non-compliant thus there is a need for the development of the oral drug delivery system which will provide improved bioavailability. Thiolatedpoly (acrylic acid) was used as a carrier to deliver the LMW heparin orally. It was observed that formulating low molecular weight heparin with thiolated polymers have shown to provide relatively improved bioavailability ($5.8 \pm 1.4\%$) [9]. LMW heparin has shown absolute bioavailability of $\geq 20\%$ in rats[34].

Transmucosal delivery of drug

Thiomers have the potential to deliver the drugs through transmucosal routes comparatively with better effects. Thiomers have the ability to provide the control drug release. In a study thiolated chitosan was prepared using thio butyl amidine as a conjugating agent. Chitosan-thio-butyl-amidine microspheres were prepared to evaluate the controlled delivery of the actives. The results have showed the controlled release of drug through these microspheres. Thus thiomers can be effectively used for the less absorbed drugs to be delivered transmucosally[35,36]. It is also concluded that the thiolated chitosan can be a good candidate for the formulation as beads for the controlled release of the drug[19].

Gastrointestinal delivery of drug

The recent advance in the polymer technology has led to improve in the drug delivery. The hydrophilic macromolecular drugs are now can be effectively delivered using thiomers due increased contact time by mucoadhesion and also due to enzyme inhibiting properties of thiomers as in case of GI peptidase[37].

Peptides;Peptide delivery can be enhanced by incorporating it with thiomers. The oxidation of peptide is reduced and also the permeation of drug through the gut is increased due to increased stay of the formulation in the body. Thiolated chitosan can enhance the stability and dissolution profile of the orally delivered peptides. One of the formulation for the oral delivery of peptides is by using the thiolated chitosan .i.e. chitosan-thio-butyl-amidine. It is compressed with tablets to deliver in gut. These tablets are coated with triglycerides to prevent adhesion in oral cavity or esophagus. In stated study enhanced bioavailability as well as stability was observed [38].

Buccal delivery of drug

Mucoadhesive buccal formulations are getting more and more intentions. Thiolated polymers can be effectively used to enhance the buccal delivery of drugs especially the drugs with problems

through trans GIT delivery of the drugs. L-cysteine conjugates were used to prepare the matrix tablets and there after the unmodified and modified polymers used in preparation are further studied for effect on mucoadhesion. Thiomers thus prove to be an effective excipient for buccal delivery of the drugs[39].

Nasal delivery of drug

Thiomers can also be effective carrier for the delivery of drug through nasal route. Enhanced mucoadhesion and permeation make the thiomers effective tool for the delivery of drug through this route. Nasal route has been shown to be effective for delivery of drug using thiomers as permeation enhancement effect is more prominent. Various thiolated polymers including chitosan, sodium alginates and polycarbophil can be used for the nasal delivery of the drug. Thiolated polycarbophil was used to study the leu-enkephalin delivery on the bovine mucosal cells. The results showed to deliver the drug through the thiomers by sustained release and the degradation of drug is lowered. Also there was increase in drug uptake by nasal mucosa by 80 folds. Thus thiolated polycarbophil can prove to be a promising tool for nasal delivery drug[35].

Human growth hormone (hGH); Thiolated polycarbophil and glutathione was used for the delivery of hGH through nasal route in the form of hydrogel. It was observed that this formulation has effectively improve the plasma level of the drug [9].

Ophthalmic delivery of drug

Ophthalmic drug delivery is an effective route of administration but it also various drawbacks and the important one is decreased retention time of the drug. Various thiomers including thiolated chitosan, thiolated sodium alginate, thiolated poly ethylene glycol and polycarbophil etc. has the potential to deliver the drug effectively with increased mucoadhesion. A thiomers of non-ionic surfactant i.e. cysteine-PEG was used to formulate the nanoparticulate preparation of the cyclosporine to deliver through the ophthalmic route and was shown to remain in the ocular cul-de-sac for about 6h. Thus it has shown to increase the stay and also the concentration of drug in ocular region as compared to normal lipid carriers[40]. Similarly ocular inserts of using thiomers are carrier like thiolated polyacrylic acid can are also be formulated to improve the bioavailability as well as effectiveness of the drugs [35]. Various studies showed that plasmid DNA encoded with green fluorescence protein can be delivered with improved effects in a study, it was nanocomplexed with modified and unmodified chitosan to compare the effects of both carrier. Thereafter the transfection efficiency of thiolated chitosan and the sustained action produced by the thiolated chitosan was found greater comparatively [35,41].

Vaginal delivery of drug

Delivery of drug through intra-vaginal route using thiomers offer various advantages like increased in-situ gelling, controlled release, mucoadhesion, enzyme inhibition etc. thereby increasing the concentration of drug delivered. Thiomeric formulation to be administered

through vaginal route can be in the form of tablets, capsules, gels, liquids etc and once administered they remain there to deliver drug at a controlled release rate for even weeks[35].

In a study thiolated carbopol was used for vaginal delivery of LH-RH to observe the effect of thiomers on aminopeptidase N. it was observed that enzyme inhibition is greatly linked with thiol concentration. Increase in the concentration of thiol group improves the enzyme inhibition[42].

Conclusion

The improvement in the mucoadhesive properties of the polymers by incorporation of the thiol groups provide us with the polymers and ultimately, the dosage form with multiple additive properties such as taste masking, improved permeation, prolonged release, reduced irritation and patient compliance. Different polymers that are modified by addition of thiol groups, are used in different preparations for different routes accordingly. By using these modified polymers the plasma drug level are increased.

References

1. Roy, S., et al., *Polymers in mucoadhesive drug-delivery systems: a brief note*. Designed monomers and polymers, 2009. **12**(6): p. 483-495.
2. Deepak, K., M.S. Kumar, and N. Mahadevan, *Thiolated chitosan: Modified advanced generation of mucoadhesive polymers*. International Journal of Recent Advance in Pharmaceutical Research, 2012. **2**(3): p. 31-41.
3. Shahnaz, G., et al., *Synthesis, characterization, mucoadhesion and biocompatibility of thiolated carboxymethyl dextran–cysteine conjugate*. Journal of Controlled Release, 2010. **144**(1): p. 32-38.
4. Anitha, A., et al., *Development of mucoadhesive thiolated chitosan nanoparticles for biomedical applications*. Carbohydrate Polymers, 2011. **83**(1): p. 66-73.
5. Mahajan, H.S., et al., *Thiolated xyloglucan: Synthesis, characterization and evaluation as mucoadhesive in situ gelling agent*. Carbohydrate polymers, 2013. **91**(2): p. 618-625.
6. Albrecht, K. and A. Bernkop-Schnürch, *Thiomers: forms, functions and applications to nanomedicine*. 2007.
7. Mythri, G., et al., *Novel Mucoadhesive Polymers—A Review*. 2011.
8. Wagh, M., et al., *Thiomers: a new generation of mucoadhesive polymers*. Research J. Pharm. and Tech, 2009. **2**(2): p. 250-255.
9. Kafedjiiski, K. and L. FRANZENS, *Multifunctional polymeric excipients in no-invasive delivery of hydrophilic macromolecular drugs: the thiomers-technology*. The drug delivery companies report Autumn/winter, 2004. **47**.
10. Kushawaha, S., M. Bansal, and P. Sharma, *Thiomers—A new generation mucoadhesive*.
11. Bhalekar, M., S. Sonawane, and S. Shimpi, *Synthesis and characterization of a cysteine xyloglucan conjugate as mucoadhesive polymer*. Brazilian Journal of Pharmaceutical Sciences, 2013. **49**(2): p. 285-292.
12. Kast, C.E. and A. Bernkop-Schnürch, *Polymer–cysteamine conjugates: new mucoadhesive excipients for drug delivery?* International journal of pharmaceutics, 2002. **234**(1): p. 91-99.
13. Biličić, M.B., et al., *Synthesis and characterization of thiomers of polyaspartamide type*. International journal of pharmaceutics, 2005. **291**(1): p. 211-219.

14. Barbarić, M., et al., *Synthesis and in vitro antitumor effect of diclofenac and fenoprofen thiolated and nonthiolated polyaspartamide-drug conjugates*. European journal of medicinal chemistry, 2007. **42**(1): p. 20-29.
15. Rahmat, D., et al., *Design and synthesis of a novel cationic thiolated polymer*. International journal of pharmaceutics, 2011. **411**(1): p. 10-17.
16. Jayakumar, R., R. Reis, and J. Mano, *Synthesis and characterization of pH-sensitive thiol-containing chitosan beads for controlled drug delivery applications*. Drug Delivery, 2007. **14**(1): p. 9-17.
17. Chopra, S., et al., *Advances and potential applications of chitosan derivatives as mucoadhesive biomaterials in modern drug delivery*. Journal of pharmacy and pharmacology, 2006. **58**(8): p. 1021-1032.
18. Zhu, X., et al., *Synthesis of thiolated chitosan and preparation nanoparticles with sodium alginate for ocular drug delivery*. Molecular vision, 2012. **18**: p. 1973.
19. Kast, C.E., et al., *Chitosan-thioglycolic acid conjugate: a new scaffold material for tissue engineering?* International journal of pharmaceutics, 2003. **256**(1): p. 183-189.
20. Hauptstein, S. and A. Bernkop-Schnürch, *Thiomers and thioemer-based nanoparticles in protein and DNA drug delivery*. Expert opinion on drug delivery, 2012. **9**(9): p. 1069-1081.
21. Bravo-Osuna, I., et al., *Mucoadhesion mechanism of chitosan and thiolated chitosan-poly (isobutyl cyanoacrylate) core-shell nanoparticles*. Biomaterials, 2007. **28**(13): p. 2233-2243.
22. Kafedjiiski, K., et al., *Synthesis and in vitro evaluation of a novel thiolated chitosan*. Biomaterials, 2005. **26**(7): p. 819-826.
23. Grabovac, V., D. Guggi, and A. Bernkop-Schnürch, *Comparison of the mucoadhesive properties of various polymers*. Advanced drug delivery reviews, 2005. **57**(11): p. 1713-1723.
24. Kast, C.E. and A. Bernkop-Schnürch, *Thiolated polymers—thiomers: development and in vitro evaluation of chitosan—thioglycolic acid conjugates*. Biomaterials, 2001. **22**(17): p. 2345-2352.
25. Wang, X., et al., *Preactivated thiomers: permeation enhancing properties*. International journal of pharmaceutics, 2012. **438**(1): p. 217-224.

26. Hornof, M.D., C.E. Kast, and A. Bernkop-Schnürch, *In vitro evaluation of the viscoelastic properties of chitosan–thioglycolic acid conjugates*. European journal of pharmaceutics and biopharmaceutics, 2003. **55**(2): p. 185-190.
27. Albrecht, K., et al., *Preparation of thiomers microparticles and in vitro evaluation of parameters influencing their mucoadhesive properties*. Drug development and industrial pharmacy, 2006. **32**(10): p. 1149-1157.
28. Rahmat, D., et al., *Thiolated hydroxyethyl cellulose: design and in vitro evaluation of mucoadhesive and permeation enhancing nanoparticles*. European Journal of Pharmaceutics and Biopharmaceutics, 2013. **83**(2): p. 149-155.
29. Davidovich-Pinhas, M., O. Harari, and H. Bianco-Peled, *Evaluating the mucoadhesive properties of drug delivery systems based on hydrated thiolated alginate*. Journal of Controlled Release, 2009. **136**(1): p. 38-44.
30. Iqbal, J., et al., *Preactivated thiomers as mucoadhesive polymers for drug delivery*. Biomaterials, 2012. **33**(5): p. 1528-1535.
31. Leitner, V.M., G.F. Walker, and A. Bernkop-Schnürch, *Thiolated polymers: evidence for the formation of disulphide bonds with mucus glycoproteins*. European Journal of Pharmaceutics and Biopharmaceutics, 2003. **56**(2): p. 207-214.
32. Bernkop-Schnürch, A., *Mucoadhesive systems in oral drug delivery*. Drug discovery today: Technologies, 2005. **2**(1): p. 83-87.
33. Guggi, D., C.E. Kast, and A. Bernkop-Schnürch, *In vivo evaluation of an oral salmon calcitonin-delivery system based on a thiolated chitosan carrier matrix*. Pharmaceutical research, 2003. **20**(12): p. 1989-1994.
34. Bernkop-Schnürch, A., M.H. Hoffer, and K. Kafedjiiski, *Thiomers for oral delivery of hydrophilic macromolecular drugs*. Expert opinion on drug delivery, 2004. **1**(1): p. 87-98.
35. Bhargav, E., *THIOMERS FRESH DRIFT OF POLYMERS & THEIR PROSPECTIVE IN PHARMACEUTICALS: A REVIEW*.
36. Elhassan Imam, M. and A. Bernkop-Schnürch, *Controlled drug delivery systems based on thiolated chitosan microspheres*. Drug development and industrial pharmacy, 2005. **31**(6): p. 557-565.

37. Bernkop-Schnürch, A., et al., *Thiomers: potential excipients for non-invasive peptide delivery systems*. European journal of pharmaceutics and biopharmaceutics, 2004. **58**(2): p. 253-263.
38. Langoth, N., J. Kalbe, and A. Bernkop-Schnürch, *Development of buccal drug delivery systems based on a thiolated polymer*. International journal of pharmaceutics, 2003. **252**(1): p. 141-148
39. Shen, J., et al., *Mucoadhesive effect of thiolated PEG stearate and its modified NLC for ocular drug delivery*. Journal of controlled release, 2009. **137**(3): p. 217-223.
40. Schmitz, T., et al., *Synthesis and characterization of a chitosan-N-acetyl cysteine conjugate*. International journal of pharmaceutics, 2008. **347**(1): p. 79-85.
41. Schmitz, T., et al., *Development and in vitro evaluation of a thiommer-based nanoparticulate gene delivery system*. Biomaterials, 2007. **28**(3): p. 524-531.
42. Valenta, C., et al., *Evaluation of the inhibition effect of thiolated poly (acrylates) on vaginal membrane bound aminopeptidase N and release of the model drug LH-RH*. Journal of pharmacy and pharmacology, 2002. **54**(5): p. 603-610.
43. Grabovac, V., D. Guggi, and A. Bernkop-Schnürch, *Comparison of the mucoadhesive properties of various polymers*. Advanced drug delivery reviews, 2005. **57**(11): p. 1713-1723.
44. Hrsic, E., H. Keul, and M. Möller, *Synthesis of thiol functionalized poly (meth) acrylates through enzymatic catalysis and a subsequent one pot reaction process*. European Polymer Journal, 2012. **48**(4): p. 761-768.
45. Kotal, A., et al., *Synthesis of semitelechelic POSS-polymethacrylate hybrids by thiol-mediated controlled radical polymerization with unusual thermal behaviors*. Journal of Polymer Science Part A: Polymer Chemistry, 2008. **46**(3): p. 1111-1123.
46. Andrade, A.O., M.E. Parente, and G. Ares, *Screening of mucoadhesive vaginal gel formulations*. Brazilian Journal of Pharmaceutical Sciences, 2014. **50**(4): p. 931-941.
47. Guggi, D., C.E. Kast, and A. Bernkop-Schnürch, *In vivo evaluation of an oral salmon calcitonin-delivery system based on a thiolated chitosan carrier matrix*. Pharmaceutical research, 2003. **20**(12): p. 1989-1994.

48. Kafedjiiski, K., et al., *Synthesis and in vitro evaluation of thiolated hyaluronic acid for mucoadhesive drug delivery*. International journal of pharmaceutics, 2007. **343**(1): p. 48-58.
49. Yang, G., G.D. Prestwich, and B.K. Mann, *Thiolated carboxymethyl-hyaluronic-acid-based biomaterials enhance wound healing in rats, dogs, and horses*. ISRN veterinary science, 2012. **2011**.
50. Southan, A., et al., *Side chain thiol-functionalized poly (ethylene glycol) by post-polymerization modification of hydroxyl groups: synthesis, crosslinking and inkjet printing*. Polymer Chemistry, 2014. **5**(18): p. 5350-5359.
51. Williams, S.R., et al., *Synthesis and Characterization of Poly (ethylene glycol)–Glutathione Conjugate Self-Assembled Nanoparticles for Antioxidant Delivery*. Biomacromolecules, 2008. **10**(1): p. 155-161.